MASS SPECTROMETRY OF PERTRIMETHYLSILYL NEURAMINIC ACID DERIVATIVES*

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ABSTRACT

The mass spectra of the trimethylsilyl (TMS) derivatives of the methyl and trideuteriomethyl esters of N-acetylneuraminic acid, the methyl ester of N-glycolylneuraminic acid, the methyl ester trideuteriomethyl β -glycoside of N-acetylneuraminic acid, the trideuteriomethyl ester trideuteriomethyl β -glycoside of N-acetylneuraminic acid, and the methyl esters of the $(2\rightarrow 3)$ - and $(2\rightarrow 6)$ -linked isomers of N-acetylneuraminic acid—lactose are discussed The characteristic fragmentation patterns of the sialic acid derivatives can be used for the identification of this type of carbohydrate The $(2\rightarrow 3)$ - and $(2\rightarrow 6)$ -linked isomers of N-acetylneuraminic acid—lactose can be differentiated

INTRODUCTION

Previously, we reported on the applicability of mass spectrometry of pertrimethylsilyl (TMS) derivatives of several classes of oligosaccharides, namely, aldosyl disaccharides¹, disaccharides consisting of an aldohexose and a 2-acetamido-2-deoxy-aldohexose², and di- and higher saccharides consisting of aldohexoses and hexuloses³ The importance of sialic acids in such biopolymers as glycoproteins and glycolipids prompted an extension of these studies to neuraminic acid derivatives. Sweeley et al have reported on the mass spectrometry of sialoglycosphingolipids^{4,5} and the TMS derivative of the methyl ester methyl glycoside of neuraminic acid⁶ Recently, Kochetkov et al⁷ described the mass spectrometry of peracetyl derivatives of some sialic acids. We now report on the mass spectrometry of the TMS derivatives of the methyl (1a) and trideuteriomethyl (1b) esters of N-acetylneuraminic acid, the methyl ester of N-glycolylneuraminic acid (2), the methyl ester methyl β -glycoside of N-acetylneuraminic acid (3a), the trideuteriomethyl ester trideuteriomethyl β -glycoside of N-acetylneuraminic acid (3b), and the methyl ester of N-acetylneuraminic acid- α -(2 \rightarrow 3)- (4) and α -(2 \rightarrow 6)-lactose (5)

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RESULTS AND DISCUSSION

Stalic acids In the mass spectra of 1a, 1b, 2, 3a, and 3b, the peak for the molecular ion (M) was very weak. The first, detectable fragment ion with reasonably high intensity in the high-mass range was $(M-CH_3)$. Tables I-III contain details of fragment ions given by 1, 2, and 3, based on exact mass measurements and/or relative shifts in mass.

TABLE I Interpretation of some important fragment ions, present in the mass spectra of the tms derivatives of the methyl (1a) and trideuteriomethyl (1b) esters of N-acetylneuraminic acid

m/eª		Formula	Fragment	m/e ^b	
668	(44 2)	C ₂₆ H ₅₈ NO ₉ S ₁₅	M-CH ₃	671	(32 2)
640	(61)	C25H58NO8S15	M-Ac	643	(47)
624	(27 7)	C25H58NO7S15	M-COOCH ₃	624	(20 9)
593	(115)	C24H51NO8S14	M-TMSOH	596	(9 5)
580	(5 0)		M-CH ₂ OTMS	583	(4 3)
578	(10 0)	C23H48NO8S14	668 - TMSOH (m* = 500 1)	581	(7 4)
<i>5</i> 52	(6 2)	$C_{22}H_{50}NO_{7}S_{14}$	$M-OTMS-CH_2=C=O$	555	(4 5)
534	(5 4)	$C_{22}H_{46}O_{7}S_{14}$	$M-TMSOH-NH_2Ac$	537	(3 4)
			$M-TMSOH-COOCH_3$	534	(0 8)
503	(3 1)		M-TMSOH-TMSOH	506	(2 3)
492	(7 7)		$M-CH_2-C(OH)(COOCH_3)(OTMS)$	492	(5 4)
			$M-COOCH_3-TMSOH-CH_2=C=O$		
490		$C_{20}H_{40}NO_7S_{13}$	M-CH ₂ OTMS-TMSOH	493	(14 6)
488	(3 5)		$M-CH_3-TMSOH-TMSOH$	491	(3 4)
478		$C_{19}H_{40}NO_7S_{13}$	M – CHOTMS–CH₂OTMS	481	(66 2)
462	(3 8)		$M-OTMS-CH_2=C=O-TMSOH$	465	(2 9)
449		$C_{18}H_{39}NO_6S_{13}$	$M-CH_2OTMS-CH_2=C=O-OTMS$	452	(140)
431		C ₁₈ H ₃₅ O ₆ S ₁₃	M-CH ₂ OTMS-TMSOH-NH ₂ Ac	434	` '
400		$C_{17}H_{30}NO_6S_{12}$	490 - TMSOH (m* = 326.5)		(29 3)
388		$C_{16}H_{30}NO_6S_{2}$	478 - TMSOH (m* = 3149)	391	(8 6)
376		$C_{15}H_{30}NO_6S_{12}$	M-CHOTMS-CHOTMS-CH ₂ OTMS	379	(8 1)
358		$C_{15}H_{28}NO_5Si_2$	$M-CH_2OTMS-TMSOH-TMSOH-CH_2=C=O$	361	(117)
330		C ₁₄ H ₂₈ NO ₄ S ₁₂	No reasonable structure can be proposed	333	(12 6)
317	(48 8)	$C_{13}H_{25}O_5S_{12}$	M-CHOTMS-CHOTMS-CH ₂ OTMS-NH ₂ Ac	320	(46 4)
300	(75 8)	C ₁₃ H ₂₆ NO ₃ S ₁₂	Ac-NH=CH-C(OTMS)=CH-CH=CHOTMS	300	(65 8)
298		C13H20NO5S1	M-CHOTMS-CH2OTMS-TMSOH-TMSOH		(100 0)
227		C ₁₀ H ₁₅ O ₄ S ₁	M-CHOTMS-CHOTMS- CH2OTMS-NH2Ac-TMSOH	230	` (9 9)
217	(30 0)	$C_9H_{21}O_2Sl_2$	TMSO-CH=CH-CH=OTMS	217	(32 4)
205	(18 8)	$C_8H_{21}O_2S_{12}$	TMSO=CH-CH₂OTMS	205	(18 7)
204	(18 5)	$C_8H_{20}O_2S_{12}$	TMSO=CH-CH-OTMS	204	(18 7)
186	(21 5)	$C_3H_{16}NO_2S_1$	AcNH-CH-CH-CH-OTMS	186	(25 9)
173	(16 2)	$C_7H_{15}NO_2S_1$	Acnh-Ch-Ch=OTMS	173	(21 4)

[&]quot;Compound 1a; the intensities of the ions, relative to that of m/e 298, are given in brackets "Compound 1b, the intensities of the ions, relative to that of m/e 301, are given in brackets

The ion at m/e 624 for 1a could be $(M-COOCH_3)$ or $(M-NH_2COCH_3)$, exact mass measurements support the former structure. The mass spectrum of 1b shows a distinct peak at m/e 624 $(M-COOCD_3)$ Calculation of the intensities of the isotope peaks of m/e 624 did not indicate the presence of a contribution to the peak at m/e 627 other than the isotope peak for m/e 624 Examination of the mass spectra of 2 and 3 suggests that, in addition to a $COOCH_3$ radical, a NH_2COR_3 $(R_3 = CH_3)$ or $CH_2OTMS)$ molecule can be eliminated via a McLafferty rearrangement. The mass spectrum of 2 shows a relatively intense peak at m/e 712

TABLE II

INTERPRETATION OF SOME IMPORTANT FRAGMENT IONS, PRESENT IN THE MASS SPECTRUM OF
THE TMS DERIVATIVE OF THE METHYL ESTER OF N-GLYCOLYLNEURAMINIC ACID (2)

m/e	a	Formula	Fragment
756	(23 5)	C29H66NO10S16	M-CH ₃
	(197)	C28H66NO8S16	M-COOCH ₃
681	(41)	C27H59NO9S15	M-TMSOH
668	(67)	C26H58NO9S15	M-CH ₂ OTMS
666	(4 3)	-20-38-1-94-3	756 - TMSOH (m* = 586 7)
640	(5 8)	C25H58NO8SI5	M-COCH ₂ OTMS
624	(16)		M-NH ₂ COCH ₂ OTMS
591	(2 2)		M-TMSOH-TMSOH
580	(5 4)		$M - CH_2 - C(OH)(COOCH_3)(OTMS)$
578	(9 0)	C23H48NO8S14	M-CH ₂ OTMS-TMSOH
576	(2 2)		$M-CH_3-TMSOH-TMSOH$
566	(54 7)	$C_{22}H_{48}NO_8S_{14}$	M-CHOTMS-CH2OTMS
537	(110)	C21H47NO7S14	M-COCH ₂ OTMS-CH ₂ OTMS
534	(3 2)		M-TMSOH-NH ₂ COCH ₂ OTMS
501	(3 2)		M-TMSOH-TMSOH-TMSOH
488	(27 1)	$C_{20}H_{38}NO_{7}S_{13}$	578 - TMSOH (m* = 412 0)
476	(68)	$C_{19}H_{38}NO_7Si_3$	566 - TMSOH (m* = 400 3)
464	(42)	$C_{18}H_{38}NO_{7}S_{13}$	M-CHOTMS-CHOTMS-CH2OTMS
431	(104)	$C_{18}H_{35}O_6S_{13}$	M-CH ₂ OTMS-TMSOH-NH ₂ COCH ₂ OTMS
418	(10 1)	$C_{17}H_{36}NO_5S_{13}$	No reasonable structure can be proposed
200 /	(100.0)	C II NO C	THEORY OF AUT OF COMMENT OF STREET
	(100 0)	C ₁₆ H ₃₄ NO ₄ S ₁₃	TMSOCH ₂ CO-NH=CH-C(OTMS)=CH-CH=CHOTMS
	(58 7)	C ₁₆ H ₂₈ NO ₆ S ₁₂	476-TMSOH (m* = 313 0)
374	(7 1)	C ₁₅ H ₂₈ NO ₆ S ₁₂	M-CHOTMS-CHOTMS-CH ₂ OTMS-TMSOH
317	(26 7)	$C_{13}H_{25}O_{5}Sl_{2}$	M-CHOTMS-CHOTMS-CH ₂ OTMS-NH ₂ COCH ₂ OTMS
274	(23 2)	$C_{11}H_{24}NO_3Si_2$	TMSOCH₂CONH-CH=CH-CH=OTMS
261	(13 3)	$C_{10}H_{23}NO_3S_{12}$	TMSOCH₂CONH-CH-CH=OTMS
227	(8 7)	C ₁₀ H ₁₅ O ₄ S ₁	M-CHOTMS-CHOTMS-
	()	10 113 1 2 1	CH2OTMS-NH2COCH2OTMS-TMSOH
217	(27.4)	$C_9H_{21}O_2S_{12}$	TMSO-CH=CH-CH=OTMS
205	(21 3)	$C_8H_{21}O_2S_{12}$	TMSO=CH-CH₂OTMS
204	(24 2)	$C_8H_{20}O_2S_{12}$	TMSO=CH-CH-OTMS

[&]quot;The figures in brackets are the intensities of the ions, relative to that of m/e 388

 $(M-COOCH_3)$ and a peak of low intensity at m/e 624 $(M-NH_2COCH_2OTMS)$ [I(624) I(712) = 1 12] In the mass spectrum of 3a, a peak at m/e 566 $(M-COOCH_3)$ and $M-NH_2COCH_3$) is observed, which is mainly shifted to m/e 569 in the spectrum of 3b Calculation of the intensities of the isotope peaks of m/e 569 indicated the

TABLE III

INTERPRETATION OF SOME IMPORTANT FRAGMENT IONS, PRESENT IN THE MASS SPECTRA OF
THE TMS DERIVATIVES OF THE TRIDEUTERIOMETHYL ESTER TRIDEUTERIOMETHYL GLYCOSIDE OF
N-ACETYLNEURAMINIC ACID AND OF THE METHYL ESTER METHYL GLYCOSIDE (3a)

m/e	,a	Fragment	m/e ^b
610	(29 5)	M-CH ₃	616 (28 3)
594	(1.7)	$M-OCH_3$	597 (17)
578	(2 3)	$610 - CH_3OH (m^* = 547.7)$	581 (17)
566	(22 7)	M-COOCH ₃	569 (21 9)
		$M-NH_2Ac$	572 (3 0)
535	$(13\ 5)$	M-TMSOH	541 (25 3)
522	(3 5)	$M-CH_2OTMS$	528 (3 9)
520	(4 3)	610 - TMSOH (m* = 443 3)	526 (43)
504	(3 1)	$M-OCH_3-TMSOH$	507 (3 0)
494	(5 7)	$M-OTMS-CH_2=C=O$	500 (52)
490	(21)	$M-CH_2OTMS-CH_3OH$	493 (19)
488	(17)	$M-CH_3-TMSOH-CH_3OH$	491 (15)
476	(11.5)	$535 - NH_2Ac (m^* = 423 5)$	482 (112)
		M-TMSOH-COOCH ₃	479 (1 5)
462	(2 3)	$M-OCH_3-CH_2=C=O-TMSOH$	465 (60)
445	(8 1)	M-TMSOH-TMSOH	451 (3 2)
434	(5 5)	$M-COOCH_3-CH_2=C=O-TMSOH$	437 (69)
432	(6 9)	M-CH ₂ OTMS-TMSOH	438 (15 5)
420	(70 5)	M-CHOTMS-CH2OTMS	426 (80 3)
391	(10 1)	$M-CH_2OTMS-CH_2=C=O-OTMS$	397 (12 0)
388	(8 1)	$420 - CH_3OH (m^* = 358 4)$	391 (5 2)
373	$(25\ 2)$	M-CH ₂ OTMS-TMSOH-NH ₂ Ac	379 (30 0)
355	(5 2)	M-TMSOH-TMSOH-TMSOH	361 (8 6)
342	(26 9)	432 - TMSOH (m* = 270 8)	348 (30 0)
318	(5 9)	M-CHOTMS-CHOTMS-CH2OTMS	324 (7 1)
300	(69 8)	Ac-NH=CH-C(OTMS)=CH-CH=CHOTMS	300 (81 5)
298	$(100\ 0)$	M-CHOTMS-CH2OTMS-CH3OH-TMSOH	301 (100 0)
259		M-CHOTMS-CHOTMS-CH2OTMS-NH2Ac	265 (39 5)
227	(5 5)	M-CHOTMS-CHOTMS-CH ₂ OTMS-NH ₂ Ac-CH ₃ OH	230 (129)
217	(24 4)	TMSO-CH=CH-CH=OTMS	217 (39 1)
205	(13 5)	TMSO=CH-CH2OTMS	205 (42 9)
204	(12 6)	TMSO=CH-CH-OTMS	204 (32 8)
186	(21 5)	AcNH-CH=CH-CH=OTMS	186 (57 1)
173	(8 5)	Acnh-ch-ch=otms	173 (20 6)

^aCompound 3a, the intensities of the ions, relative to that of m/e 298, are given in brackets ^bCompound 3b, the intensities of the ions, relative to that of m/e 301, are given in brackets

presence of a small peak at m/e 572, besides the isotopic contribution to this peak [I(572) I(569) = 1 17] Similarly, it can be deduced that the peak at m/e 534 in the spectrum of 1a is due mainly to (M-TMSOH-NH₂COCH₃); the spectrum of 1b contains a small peak at m/e 534 and a more intense peak at m/e 537 Π (534) Π (537) = 1 4] In the spectrum of 2, a distinct peak at m/e 534 (M-TMSOH-NH₂COCH₂-OTMS) is observable, besides a peak of low intensity at m/e 622 (M-TMSOH-COOCH₃) $\Pi(622)$ $\Pi(534) = 1.7$ The spectrum of 3a shows a peak at m/e 476, which is shifted mainly to m/e 482 but also partly to m/e 479 in the spectrum of 3b [I(479) I(482) = 1 8]. These data, together with the presence of metastable peaks at 423 5 and 429 4 in the spectra of 3a and 3b, respectively, make it clear that (M-TMSOH) eliminates mainly NH₂COCH₃, (m* = 423 5 $535^+ \rightarrow 476^+ + \text{NH}_2$ - $COCH_3$, $m^* = 429 4$ $541^+ \rightarrow 482^+ + NH_2COCH_3$), in contrast to the molecular ion which eliminates mainly COOCH₃ From the different spectra, it can be deduced that (M-COOCH₃) is hardly degraded further However, the elimination of NH₂COCH₃ (or NH₂COCH₂OTMS) via a McLafferty rearrangement occurs frequently, probably mainly from fragment ions. In Fig. 1, the formation of (M-[CHOTMS-CHOTMS-CH₂OTMS]-NH₂COR₃-R₂OH) is shown.

Fig 1 Formation of M-[CHOTMS-CHOTMS-CH₂OTMS]-NH₂COR₃-R₂OH

The neuraminic acid derivatives can eliminate CH₂OTMS (C-9), CHOTMS-CH₂OTMS (C-8,9), and CHOTMS-CHOTMS-CH₂OTMS (C-7,8,9), respectively. Because of the intensities of the peaks for (M-[CHOTMS-CH₂OTMS]) in each spectrum, the second fragmentation pathway appears to have a high preference

The peak at m/e 298, which is one of the most intense peaks in the mass spectra of 1a and 3a, is shifted 3 m u in the spectra of 1b and 3b, and 88 m u. in the spectrum of 2. In Fig 2, fragmentation schemes are given for the different compounds It can be deduced from the spectra that the C-8,9 fragment is first eliminated from the molecular ion, followed by consecutive eliminations of R_2OH ($R_2 = CH_3$, CD_3 , or TMS) in which OR_2 originates from C-2, and TMSOH in which OTMS originates from C-4. Furthermore, it can be concluded that the elimination of the C-9-fragment from M is combined with the elimination of R_2OH ($R_2 = CH_3$, CD_3 , or TMS) in which OR_2 originates from C-2, or with a TMSOH elimination in which OTMS originates from C-4 or C-7

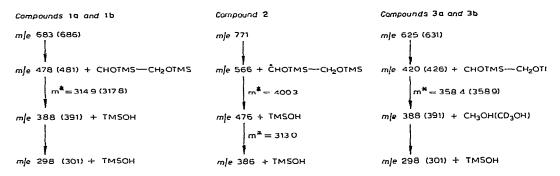


Fig 2 Formation of m/e 298 from 1a and 3a, m/e 301 from 1b and 3b, and m/e 386 from 2

Fig. 3. Formation of m/e 300 from 1 and 3, and m/e 388 from 2

The fragment ion at m/e 300, which is the second most-intense peak in the spectra of 1a, 1b, 3a, and 3b, is shifted to m/e 388 in the spectrum of 2. It does not contain the substituents at C-2 (COOR₁ with R₁ = CH₃ or CD₃, OR₂ with R₂ = CH₃,

 CD_3 , or TMS) From metastable measurements, using a defocusing technique according to Barber and Elliott, these fragment ions are identified as $\{M-TMSOH-TMSOH-[O=CH-CH_2-C(COOR_1)OR_2]\}$ A fragmentation scheme is presented in Fig. 3

The origin of the peaks at m/e 204 and 217 is quite different from that in the spectra of the TMS-aldohexoses⁸ The fragment at m/e 217 can be formed from the side chain C-7,8,9 by elimination of TMSOH from m/e 307 (CH₂OTMS-CHOTMS-CHOTMS-CH-OTMS \rightarrow TMSOCH-CH-CH-OTMS+TMSOH) The peak at m/e 204 can be formed from the side chain by loss of a CH₂OTMS radical (CH₂OTMS-CHOTMS-CH-OTMS \rightarrow TMSO-CH-CH-OTMS+CH₂OTMS) and then it comprises the fragment C-7,8 The peaks at m/e 173 and 186 in the mass spectra of 1 and 3 contain the N-acetyl group and are shifted 88 m u in the spectrum of 2 to m/e 261 and 274, respectively (N-glycolyl group) If no migrations take place, the fragment ion at m/e 173 contains C-4,5 (Ac-NH-CH-CH-OTMS) Fig 4 shows the formation of the fragment m/e 186, containing C-5,6,7 (Ac-NH-CH-CH-CH-OTMS) However, Dawson et al ⁵ suggest that this ion has the structure C-3,4,5 (Ac-NH-CH-C(OTMS)=CH₂) The formation of this ion is possible only if a hydrogen transfer from C-4 is assumed Labelling on C-4 and C-7 can discriminate between these alternatives

Fig 4 Formation of m/e 186 from 1 and 3, and m/e 274 from 2

Trisaccharides consisting of sialic acid and lactose. The mass spectra of the TMS derivatives of the methyl esters of N-acetylneuraminic acid- α -(2 \rightarrow 3)- (4) and α -(2 \rightarrow 6)-lactose (5) show a great similarity. Some significant peaks are given in Table IV. A molecular ion peak (m/e 1439) was observed in each spectrum. The peaks at m/e 1424, 1380, and 1234 are assigned to (M-CH₃), (M-COOCH₃ or NH₂-COCH₃), and (M-[CH₂OTMS-CHOTMS]), respectively. The mass spectrum of the (2 \rightarrow 6)-isomer shows a peak at m/e 726 (C₂₉H₆₄NO₁₀S1₅), which is the analogue of the peak at m/e 583 in TMS-aldohexosyl-(1 \rightarrow 6)-aldohexoses¹. The (2 \rightarrow 3)-isomer does not yield this fragment ion

TABLE IV
intensities of some significant peaks in the mass spectra of the (2 $ ightarrow3$)- and
$(2\rightarrow 6)$ -linked isomers of N-acetylneuraminic acid methyl ester–lactose in percentages
relative to the intensity of the peak at m/e 204

m/e	2→3	2→6	m/e	2→3	2→6
1439	0 6	0.7	504	31 9	13 8
1424	2 1	17	451	18	16
1380	18	18	361	15 1	7 1
1234	5 8	29	300	12 5	14 6
972	4 6	2 5	298	9 5	8 3
726	0 6	3 1	217	64 3	57 8
683	06	3 3	204	100 0	100 0
624	70	79	186	24 2	19 3
595	12 1	89	173	12 1	7 5
594	22 4	89			

In the spectrum of the $(2\rightarrow6)$ -isomer, there is also a peak at m/e 683 ($C_{27}H_{63}O_8S_{16}$) In such aldohexosylaldohexoses as lactose, the main contribution to the peak at m/e 683 stems from the reducing end of the molecule and can be attributed to TMSO-CH=C(OTMS)-CH=O-Glycose In the $(2\rightarrow3)$ -isomer, C-3 of the galactose residue is attached to the sialic acid residue. Therefore, in the latter isomer, this fragment cannot easily be formed. Each mass spectrum shows a relatively intense peak at m/e 594 ($C_{24}H_{52}NO_8S_{14}$). Moreover, in the spectrum of the $(2\rightarrow6)$ -isomer, a peak at m/e 595 ($C_{24}H_{55}O_7S_{15}$) was present in addition to the isotope peak of m/e 594. The observed peak-intensity ratio 595/594 is 0.54 for the $(2\rightarrow3)$ -isomer, and 1.00 for the $(2\rightarrow6)$ -isomer, whereas the calculated ratio is 0.49. The structure of the fragment ion with m/e 595 is TMSO-CH=CH-CH=O-Glycose. The attachment of sialic acid at C-3 of the galactose residue in the $(2\rightarrow3)$ -isomer hinders the formation of this ion

Cleavage of the glycosidic bond between C-1 of the galactose residue and the glycosidic oxygen atom, with retention of the charge at the galactose residue, leads to the fragment ion at m/e 972, and cleavage between C-2 of the neuraminic acid residue and the glycosidic oxygen atom, with retention of the charge at the neuraminic acid residue, leads to m/e 594 ($C_{24}H_{52}NO_8S_{14}$) and 504 ($C_{21}H_{42}NO_7S_{13}$, 594—TMSOH) Splitting of the glycosidic linkage between the glycosidic oxygen atom and C-4 of the glucose residue, with retention of the charge at the glucose residue, gives rise to peaks at m/e 451 and 361 ($C_{15}H_{33}O_4S_{13}$, 451—TMSOH)

Previously, we postulated that, in oligosaccharides, migration of an OTMS group occurs between saccharide units¹⁻³. For a disaccharide, a pertrimethylsilyl monosaccharide is formed as intermediate To explain the peak at m/e 624 ($C_{25}H_{58}NO_7Sl_5$), migration of an OTMS group to C-2 of the neuraminic acid residue is assumed, resulting in the formation of the TMS derivative of the methyl ester of N-acetylneuraminic acid as an intermediate from which a COOCH₃ radical is eliminated

In comparison with the spectra of the TMS derivatives of the methyl ester of N-acetylneuraminic acid and its methyl glycoside, the intensities of the peaks at m/e 300 and 298 in the spectra of the trisaccharides are strongly diminished. However, the peaks at m/e 204 and 217 are amongst the most intense peaks in the spectra, owing to the presence of the lactose molecule in the trisaccharides. The occurrence of the peaks at m/e 186 and 173 can be used for the identification of an N-acetyl group in the neuraminic acid residue

The characteristic fragmentation patterns of the TMS derivatives of the methyl esters of the N-acylneuraminic acids and their methyl glycosides indicate that mass spectrometry may be of great help in the identification of sialic acids. For oligosaccharides containing a sialic acid residue attached to lactose, it has been shown that the $(2\rightarrow 3)$ - and $(2\rightarrow 6)$ -linked isomers can be distinguished

EXPERIMENTAL

N-Acetylneuraminic acid was purchased from Calbiochem, N-glycolylneuraminic acid and N-acetylneuraminic acid- α - $(2\rightarrow 3)$ -lactose were obtained from Sigma Chemical Company, and N-acetylneuraminic acid- α - $(2\rightarrow 6)$ -lactose was a gift of Dr A Gauhe (Heidelberg) N-Glycolylneuraminic acid was contaminated with N-acetylneuraminic acid Before use, it was purified by preparative chromatography on Whatman No 3MM paper, using butyl acetate-acetic acid-water⁹ (3 2 1), detection was effected with Ehrlich's reagent¹⁰ The trideuteriomethyl ester of N-acetylneuraminic acid and its trideuteriomethyl glycoside were prepared according to procedures described by Yu and Ledeen¹¹ The methyl ester of N-glycolylneuraminic acid was prepared by the method of Sweeley et al ¹², which could also be used for the preparation of the methyl ester of N-acetylneuraminic acid. The methyl esters of the trisaccharides were obtained by the method of Derevitskaya et al ¹³ The pertrimethylsilyl derivatives of the different compounds were synthesised as described earlier¹ If necessary, the TMS derivatives were purified by preparative g1 c (Pye 105 gas chromatograph, 10% of OV-17 on Chromosorb W-AW-DMCS, 30-60 mesh)

The 70-eV mass spectra were recorded on A E I MS-9 and MS-902 mass spectrometers at an ion-chamber temperature of 80–100° for the sialic acid, and 120–140° for the trisaccharides containing sialic acid. High-resolution, mass measurements were performed at a scan speed of 16 sec per mass decade by using an A E I MS-902 mass spectrometer connected on-line with a Ferranti Argus 500 computer. The exact masses measured were converted into element lists as described by Van't Klooster et al. 14 The molecular formulae omitted from Tables I and II could not be determined because of the low intensity of the associated peaks

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REFERENCES

- 1 J P. KAMERLING, J F G VLIEGENTHART, J VINK, AND J J DE RIDDER, Tetrahedron, 27 (1971) 4275
- 2 J P KAMERLING, J F G VLIEGENTHART, J VINK, AND J J DE RIDDER, Tetrahedron, 27 (1971) 4749
- 3 J P KAMERLING, J F G VLIEGENTHART, J VINK, AND J J DE RIDDER, Tetrahedron, 28 (1972) 4375.
- 4 C C SWEELEY AND G DAWSON, Biochem Biophys Res Commun, 37 (1969) 6
- 5 G DAWSON AND C C SWEELEY, J Lipid Res, 12 (1971) 56
- 6 C C SWEELEY AND D E VANCE, Lipid Chromatogr Anal, 1 (1967) 476
- 7 N K Kochetkov, O S Chizhov, V I Kadentsev, G P Smirnova, and I G Zhukova, Carbohyd Res, 27 (1973) 5
- 8 D C DEJONGH, T RADFORD, J D HRIBAR, S HANESSIAN, M BIEBER, G DAWSON AND C C SWEELEY, J Amer Chem Soc, 91 (1969) 1728
- 9 R G SPIRO, J Biol Chem., 235 (1960) 2860
- 10 E SVENNERHOLM AND L SVENNERHOLM, Nature (London), 181 (1958) 1154
- 11 R K YU AND R LEDEEN, J Biol Chem , 244 (1969) 1306
- 12 C C SWELLEY, R BENTLEY, M MAKITA, AND W W. WELLS, J Amer Chem Soc, 85 (1963) 2479
- 13 V A DEREVITSKAYA, V M KALINEVICH, AND N K KOCHETKOV, Dokl Akad Nauk SSSR, 160 (1965) 596
- 14 H A VAN'T KLOOSTER, J S VAARKAMP-LIJNSE, AND G DIJKSTRA, Org Mass Spectrom, 8 (1974) 303